



Trabajo Original

Valoración nutricional

Cytokines and body adiposity in young female undergraduate students

Citoquinas y adiposidad corporal en estudiantes universitarias jóvenes

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Abstract

Objective: to identify cytokines and to associate them with several indexes of total and central adiposity in young female undergraduate students.

Methods: 58 young female sophomore students, aged 18 to 25 years, from a Brazilian public university were evaluated. Both anthropometric measures (weight, height, waist circumference and hip circumference) and body composition were assessed through DXA, and the values of android, gynoid and truncal fat mass were obtained. Cytokines (IL-8, IL-1 β , IL-6, IL-10 e TNF- α) were analyzed, and Body Mass Index (BMI), Body Adiposity Index (BAI), Visceral Adiposity Index (VAI), Conicity Index (CCI), Waist-Hip Index (WHR), Waist-to-Height Ratio (WHtR), Fat Mass Distribution Index 1 (FMI₁) and Fat Mass Distribution Index 2 (FMI₂) were calculated. Eventually, a linear regression was carried out to determine the regression coefficient and confidence interval (CI), having the predictor variables (cytokines) adjusted according to age and family history of obesity. The statistical significance of $\alpha = 5\%$ was applied.

Results: a correlation between adiposity indexes and cytokines (CCI, WHR and IL-12; CCI, WHR, FMI₁, FMI₂ and TNF- α) was identified. When it comes to the regression models, cytokines increase was related to CCI, WHR, FMI₁ and FMI₂ increase.

Conclusion: pro-inflammatory cytokines were associated with an increase in adipose indexes. Therefore, these indexes became a feasible strategy for clinical practice in order to identify propensity to inflammatory disorders.

Keywords:

Body fat. DXA. Anthropometry. Women.

Resumen

Objetivo: identificar citoquinas y asociarlas con los distintos índices de adiposidad total y central en estudiantes universitarias jóvenes.

Métodos: se evaluaron 58 jóvenes estudiantes, de 18 a 25 años de edad, de segundo curso de carrera de una universidad pública brasileña. Se analizaron mediante densitometría (DEXA) tanto las medidas antropométricas (peso, talla, perímetro de la cintura y perímetro de la cadera) como la composición corporal, obteniéndose los valores de masa grasa androide, ginoide y troncal. Se analizaron las citoquinas (IL-8, IL-1 β , IL-6, IL-10 y TNF- α) y se calcularon el índice de masa corporal (IMC), el índice de adiposidad corporal (IAC), el índice de adiposidad visceral (IAV), el índice de conicidad (CCI), el índice cintura-cadera (WHR), la ratio cintura-talla (WHtR), el índice de distribución de la masa grasa 1 (FMI₁) y el índice de distribución de la masa grasa 2 (FMI₂). Finalmente se realizó una regresión lineal para determinar el coeficiente de regresión y el intervalo de confianza (IC), ajustando las variables predictivas (citoquinas) a la edad y los antecedentes familiares de obesidad. Se aplicó una significación estadística de $\alpha = 5\%$.

Resultados: se detectó una correlación entre índices adiposos y citoquinas (CCI, WHR e IL-12; CCI, WHR, FMI₁, FMI₂ and TNF- α). Conforme a los modelos de regresión, el aumento de las citoquinas se relacionó con el aumento de CCI, WHR, FMI₁ y FMI₂.

Conclusión: las citoquinas proinflamatorias se asociaron al aumento de los índices adiposos. Por tanto, los índices se convierten en una estrategia factible para detectar la propensión hacia los trastornos inflamatorios en la práctica clínica.

Palabras clave:

Grasa corporal. DEXA. Antropometría. Mujeres.

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All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. The study was approved by the Comitê de Ética em Pesquisa com Seres Humanos, Universidade Federal de Viçosa (CAAE: 53452916.3.0000.5153).

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INTRODUCTION

Overweight is a public health issue related to non-communicable diseases (NCDs) both in developed and developing countries (1). This is due to the fact that obesity is a risk factor for the emergence of diseases, including hypertension, type-2 diabetes, and cardiovascular diseases. Therefore, the obesity epidemic must be the main focus of health policies, programs and actions. Thus, it is essential to implement preventive strategies such as early identification of individuals at higher risk, which may generate lower healthcare expenditures (2).

One of the strategies for early diagnosis is performing a body composition assessment, enabling the detection of potential risk to health. This assessment contributes to identify body fat accumulation even in individuals with an appropriate body weight (3). Another harmful factor related to weight gain is increased visceral adipose tissue (VAT) (4).

Increased body fat is related to the expression of the adipokines tumor necrosis factor (TNF- α) and interleukin 6 (IL-6), which contribute to the emergence of diseases (5). However, it is not always possible to measure cytokines in professional practice. One of the strategies that can be applied to assess body fat accumulation is an anthropometric assessment. This is stated based on researches (6-8) that show an association between anthropometric measures, VAT, and risk of diseases, but additive and interactive effects with visceral adiposity based on the use of cytokine indexes are still poorly studied.

A risk group for changes in VAT are undergraduate students, since they are predisposed to metabolic changes resulting from excessive visceral and body fat. This can be justified by lifestyle changes driven by joining the university, which may instigate low levels of physical activity and higher consumption of calorie-rich fast-food and ultra-processed foods (9-12). Among students, women constitute a group that physiologically may have higher fat mass and lower fat-free mass, as men have higher body weight and waist circumference values (13). These are key features to develop disease risk factors (7). These changes, triggered by excessive body fat, also increase circulating cytokines such as IL-6, C-reactive protein (CRP) and TNF- α (14), which promote the occurrence of atherosclerosis, hypertension, insulin resistance, dyslipidemia, and lipid profile changes (15,16).

Therefore, this paper aims to identify these cytokines and associate them with distinct indexes of total and central body adiposity in young female undergraduate students.

METHODS

SAMPLE

The current research is the baseline study for the Project "*Efeito do exercício físico no controle metabólico, marcadores inflamatórios, adipocinas e microbiota intestinal*" (Physical activity effects on metabolic control, inflammatory markers, adipokines, and gut microbiota). This project was carried out with a sample that con-

sisted of 75 volunteer female sophomore students, aged 18 to 25 years, from a Brazilian public university.

In order to assess the selection criteria, we evaluated sedentary or insufficiently active subjects (17) with "regular" menstrual cycles and without impairments (physical, intellectual, visual, hearing). Pregnant women, mothers with children up to 6 months of age, pacemaker users, and women taking psychotropic medication, undergoing nutritional follow-up, or suffering from diabetes or hypertension were excluded.

The sample frame of the current study consisted of 58 female undergraduate students who had available data related to cytokines.

ETHICAL PROCEDURES

The current study was approved by the Ethics Committee at *Universidade Federal de Viçosa – UFV*, protocol number CAAE: 53452916.3.0000.5153. All procedures were carried out according to the Guidelines Regulating Research Involving Human-Beings (Resolution 466/2012 of the National Health Council). All the individuals participating in it signed an informed consent form in accordance with the Declaration of Helsinki from 1975, revised in 1983.

ANTHROPOMETRIC INDEXES

Body weight was measured in kilograms by deploying a Kratoscas[®] scale. Height was measured in centimeters using a vertical portable stadiometer (Altuxata[®], Belo Horizonte, Brazil). Waist circumference (over the umbilical scar and at the midpoint between the lowest rib and the iliac crest) and hip circumference were measured using a flexible non-elastic measuring tape following the International Society for the Advancement of Kinanthropometry (ISAK) recommendations. Body adiposity index (BAI) (18), visceral adiposity index (VAI) (8), conicity index (CCI) (19), waist-hip index (WHR) (20), waist-to-height ratio (WHtR) (21) and body mass index (BMI) (20) were calculated (Table I).

BODY COMPOSITION

Body composition was assessed from 7 a.m. to 9:30 a.m. using dual energy X-ray absorptiometry (DXA) (Lunar Prodigy Advance DXA System[®] – analysis version: 13.31, GE Healthcare, Madison, WI, USA). Data regarding total body fat (TBF), regional fat (android and gynoid), lean body mass, and free fat mass were obtained. The subjects fasted for 12 hours and followed the DXA manufacturer recommendations before performing this procedure.

The android region was measured around the waist, between the midpoints of the lumbar spine and the upper part of the pelvis, whereas the gynoid region was measured approximately between the femoral head and mid-thigh (hip). Before each scan session, the device was calibrated according to the pattern procedures recommended by the manufacturer.

The DXA data obtained were used to calculate the fat mass distribution index 1 (FMI_1), mass distribution index 2 (FMI_2), and android-gynoid ratio (AGR) (Table II).

The undergraduate students who presented AGR results ≥ 1 were identified as android, while the ones with AGR results < 1 were identified as gynoid.

INFLAMMATORY CYTOKINES, CHOLESTEROL AND FRACTION

Blood samples were collected from 7 a.m. to 9 a.m., after a 12-hour fasting period, by trained professionals in an accredited laboratory. They collected 4 mL of blood from the cubital vein, which

were later centrifuged in order to separate the serum from the other blood components. Total cholesterol, low density lipoproteins (LDL), high density lipoproteins (HDL) and triglycerides (TGL) were measured.

Plasma was stored at $-80\text{ }^{\circ}\text{C}$. Then, cytokines IL-8, IL-1 β , IL-6, IL-10 and TNF- α were measured using the Cytometric Bead Array (CBA) Kit for inflammatory cytokines (INFLAMMATORY CYTOKINES CBA KIT, BD, Pharmingen, USA; Catalog no. 551811). The CBA Kit merges together the technologies of ELISA and flow cytometry, which uses polystyrene balls labeled with fluorescence at several levels (22). The data obtained from flow cytometry (FACScalibur, BD, USA) were analyzed using a specific software for CBA (FCAP Array TM Software, BD, Pharmingen, USA) through calibration curves resulting from the kit's cytokine patterns. After that, the concentration of analytes in the sample was determined in $\mu\text{g/mL}$.

Table I. Adiposity indexes and their formulas

Indexes	Formulas
Body Adiposity Index	$BAI = \frac{HC\text{ (cm)}}{\text{height (m)} \times \sqrt{\text{height (m)}}} - 18$
Visceral Adiposity Index	$VAI = \left(\frac{WC\text{ (cm)}}{36.58} + (1.89 \times BMI) \right) \times \frac{TGL}{0.81} \times \frac{1.52}{HDL}$
Conicity Index	$CCI = \frac{WC\text{ (cm)}}{0.109 \sqrt{\frac{\text{weight (kg)}}{\text{height (m)}}}}$
Waist-Hip Ratio	$WHR = \frac{WC\text{ (cm)}}{HC\text{ (cm)}}$
Waist-to-Height Ratio	$WHtR = \frac{WC\text{ (cm)}}{\text{height (m)}}$
Body Mass Index	$BMI = \frac{\text{weight (kg)}}{\text{height (m)}^2}$

HC: hip circumference (cm); WC: waist circumference (cm); BMI: body mass index (kg/m^2); TGL: triglycerides (mg/dL); HDL: high density lipoprotein (mg/dL).

Table II. Adiposity indexes, calculated from DXA results, and their formulas

Indexes	Formulas
Fat Mass Distribution Index 1	$FMI_1 = \frac{\%FT}{\%FL}$
Fat Mass Distribution Index 2	$FMI_2 = \frac{FMT}{FML}$
Android-Gynoid Ratio	$AGR = \frac{\%AF}{\%GF}$

FMI_1 : fat mass distribution index 1; FT: percentage of fat in the torso; FL: percentage of fat in the legs; FMI_2 : fat mass distribution index 2; FMT: fat mass in the torso; FML: fat mass in the limbs (fat mass in arms and legs); AGR: android-gynoid ratio; AF: android fat; GF: gynoid fat.

STATISTICAL ANALYSES

The statistical analyses were carried out with the statistical software Stata (version 13). Furthermore, a Shapiro-Wilk test was applied to check the normality of quantitative data. Their descriptive analysis is presented as median and interquartile interval.

Spearman’s correlation coefficients were applied to check the correlation between the indexes of central and body adiposity (BMI, BAI, VAI, CCI, WHtR, WC, WHR, AGR, FMI₁ and FMI₂) and the study cytokines (IL-1β, IL-6, IL-8, IL-10, IL-12 and TNF-α).

In a bivariate analysis, the regression coefficient and the confidence interval (CI) were estimated through linear regression, with adjusted variables at p ≤ 0.25 for inclusion in the model. The level of significance applied was α = 5 %. The White test was applied to identify heteroscedasticity in the distribution of errors in order to verify the adequacy of the linear regression model.

RESULTS

Fifty-eight female undergraduate students aged 18 to 25 years (± 1.82) were assessed. According to the body composition assessment, the average percentage (%) of total fat resulting from DXA was 33.61 % (± 0.94), whereas the ratio percentage of android fat to gynoid fat was 0.61 % (± 0.02). These data show that body fat is located mostly in the hip area (gluteofemoral region and thighs) (Table III).

It was seen that all adiposity indexes, except VAI, were correlated with cytokines, which were identified as weak. It is important to highlight that IL-1β did not correlate with any of the indexes, while TNF-α correlated with 70 % of them (Table IV).

Table V presents the final models of the multiple linear regression analysis; increased interleukins relate to the increased indexes CCI, WHR, FMI₁ and FMI₂, regardless of age and family history of obesity.

Table III. Physical and metabolic features of young female undergraduate students

Variable	Average	p25 – p75
Anthropometrics and body composition		
Body mass (kg)	58.39	51.39-65.39
Lean body mass (kg)	35.76	32.50-38.85
Fat mass (kg)	20.24	15.04-24.23
Bone mass (kg)	2.24	1.99-2.51
Fat-free mass (kg)	38.02	34.37-41.37
Android fat mass (kg)	0.87	0.46-1.25
Gynoid fat mass (kg)	4.01	2.96-4.55
Waist circumference – umbilical scar (cm)	76.38	69.38-83.75
Waist circumference – midpoint (cm)	76.88	70.31-3.38
Hip circumference	97.88	90.88-101.38
% of total fat from DXA	34.09	28.59-38.17
Indexes		
% Android-Gynoid Fat Ratio	0.64	0.48-0.75
BMI (kg/m ²)	22.27	19.29-24.50
WHR (cm)	0.79	0.75-0.84
WHtR (cm)	0.47	0.44-0.52
Conicity Index	1.18	1.14-1.22
Body Adiposity Index	28.43	26.50-32.08
Visceral Adiposity Index	2.40	1.93-3.53
Biochemical parameters		
Total cholesterol (mg/dL)	153.00	140.00-173.00
LDL (mg/dL)	82.00	65.00-97.00
HDL (mg/dL)	55.00	51.00-62.00
TGL (mg/dL)	76.00	60.00-111.00
Cytokines		
IL-1β (pg/mL)	123.00	65.20-236.30
IL-6 (pg/mL)	129.20	89.10-186.00
IL-8 (pg/mL)	138.40	58.30-264.50
IL-10 (pg/mL)	97.90	73.60-118.30
IL-12 (pg/mL)	131.90	62.80-249.30
TNF-α (pg/mL)	178.00	27.70-300.90

DXA: dual-energy X-ray absorptiometry; BMI: body mass index; WHR: waist-hip ratio; WHtR: waist-to-height ratio; LDL: low density lipoproteins; HDL: high density lipoproteins; TGL: triglycerides; IL: interleukin; TNF-α: tumor necrosis factor.

Table IV. Correlation between interleukins and total and central adiposity indices in young female undergraduate students

Variables	IL-1 β		IL-6		IL-8		IL-10		IL-12		TNF- α	
	R	p	R	P	r	p	R	p	r	P	r	p
BMI	-0.0646	0.6301	0.0291	0.8281	-0.1784	0.1802	-0.2340	0.0771	0.0011	0.9934	0.0397	0.7673
BAI	0.0425	0.7512	0.0531	0.6923	-0.0641	0.6327	-0.2216	0.0946	-0.0217	0.8718	-0.0140	0.9170
VAI	0.0500	0.7092	0.1234	0.3559	0.0410	0.7601	-0.1362	0.3081	0.1149	0.3906	0.0660	0.6226
CCI	0.1415	0.2894	0.2541	0.0543	0.1220	0.3616	0.0263	0.8447	0.3654	0.0048	0.3957	0.0021
WHR	0.0012	0.9931	0.1074	0.4223	-0.0941	0.4823	-0.1611	0.2269	0.1195	0.3716	0.1757	0.1872
WC	-0.0075	0.9556	0.1104	0.4094	-0.0862	0.5199	-0.1771	0.1836	0.1553	0.2245	0.1957	0.1411
WHR	0.0373	0.7809	0.1750	0.1890	0.0025	0.9850	0.0411	0.7594	0.2671	0.0427	0.3198	0.0144
AGR	0.0694	0.6045	0.1497	0.2622	-0.0402	0.7646	-0.0700	0.6017	0.1615	0.2258	0.2212	0.0953
FMI ₁	0.0594	0.6576	0.1701	0.2019	-0.0038	0.9775	-0.0035	0.9790	0.1942	0.1441	0.2666	0.0430
FMI ₂	0.0538	0.6885	0.1627	0.2224	-0.0067	0.9603	-0.0191	0.8867	0.1870	0.1599	0.2722	0.0388

BMI: body mass index; BAI: body adiposity index; VAI: vascular adiposity index; CCI: conicity index; WHR: waist-to-height ratio; WC: waist circumference measured over the umbilical scar; WHR: waist-hip ratio; AGR: android-gynoid ratio; FMI₁: fat mass distribution index 1; FMI₂: fat mass distribution index 2; IL: interleukin; TNF- α : tumor necrosis factor.

The other indexes did not meet the assumptions of the linear regression, that is, the homoscedasticity and normality of the residuals.

DISCUSSION

The higher prevalence of overweight and obesity is a fact in several countries worldwide, including Brazil. This body fat accumulation is a risk factor for diseases such as cardiovascular conditions, type-2 diabetes and hypertension (23). The sample assessed showed that 46.70 % of subjects had a high percentage of body fat. When it comes to women, a body fat accumulation higher than expected is associated with risk for developing cardiometabolic diseases (5).

Another factor that deserves attention is body fat location, for the pattern of body fat distribution in women is related to estrogen levels, which tend to decrease over the years. It causes changes regarding where fat is stored, increasing the android region and, consequently, the risk of developing diseases (24). This implies that the risks for health are related to where body fat is stored. A higher level of android fat and/or its proportion in relation to gynoid fat, for instance, is linked to risk factors for developing diseases (25).

It is known that women tend to have a higher quantity of body fat than men. Most of the fat in a woman's body is located in the peripheral region (gynoid), while it is mostly found in the abdominal region in men (android) (26). This fact confirms the results found by the current study (Table III). Although gynoid fat is less associated with cardiometabolic risks when compared to android fat (27), 35.0 % of the sample had changes in waist circumference, which predispose to dyslipidemia (28).

Adipose tissue, mainly found in the central region of the body, has a significant role in the production of inflammatory cytokines. This capacity is even greater in intra-abdominal (visceral) adipose tissue when compared to subcutaneous adipose tissue (29).

Visceral fat may produce higher levels of IL-6 and TNF- α in people with central obesity: that is, increased body fat, which may be identified by body fat indexes, can result in increased cytokine production (30).

Inflammatory parameters are indexes related to the risk of developing NCDs because they result from molecule secretion with inflammatory features, which is stimulated by the adipose tissue (31). This chronic inflammation tends to be triggered by excessive body fat, which increases circulating cytokines such as IL-6 and TNF- α (14).

The concentrations of IL-6 and TNF- α are directly proportional to the amount of adipose tissue (32). TNF- α is responsible for endothelial changes, phagocyte oxidative metabolism stimuli, and increased adipocyte activity. IL-6, produced by monocytes and endothelial cells, is responsible for increased levels of CRP, which is a marker of inflammation (15,16).

IL-6 can act in several ways in the body according to its concentration, and plays a role in the production of acute phase proteins, which are related to post-inflammatory response. Furthermore, its increase can be directly associated with body mass and insulin resistance, thus being related to subclinical inflammation in obesity (33).

TNF- α presents higher serum concentrations in individuals with high VAT in comparison to those without central obesity (34). This may be confirmed by the results found in this study (Table IV).

IL-12 plays a role in cell-mediated immunity, and is necessary to provide resistance to intracellular infection. Therefore, this is a cytokine that can be key in the development of autoimmunity, although conflicting data have been reported about it (35). The plasma concentrations of this cytokine are higher in individuals suffering from overweight or obesity in comparison to eutrophic ones. Hence, IL-12 correlates positively with fat mass.

A possible explanation for these high levels stems from the presence of hyperleptinemia in individuals with body fat changes (36). According to Fantuzzi (37), leptin favors the production

Table V. Final models of the multiple linear regression analysis between body adiposity indexes (dependent variables) and cytokines in young female undergraduate students

Variables	adjusted β^*	95 % CI	p-value
CCI			
IL-12	9.490×10^{-5}	$6.440 \times 10^{-5} - 1.835 \times 10^{-4}$	0.036
WHR			
TNF- α	6.960×10^{-5}	$1.270 \times 10^{-5} - 1.265 \times 10^{-4}$	0.017
FMI₁			
IL-6	5.002×10^{-4}	$7.530 \times 10^{-5} - 9.251 \times 10^{-4}$	0.022
IL-12	2.774×10^{-4}	$5.510 \times 10^{-5} - 4.996 \times 10^{-4}$	0.015
TNF- α	2.980×10^{-4}	$1.193 \times 10^{-4} - 4.768 \times 10^{-4}$	0.002
FMI₂			
IL-6	3.994×10^{-4}	$6.770 \times 10^{-5} - 7.310 \times 10^{-4}$	0.019
IL-12	2.226×10^{-4}	$4.930 \times 10^{-5} - 3.960 \times 10^{-4}$	0.013
TNF- α	2.330×10^{-4}	$9.320 \times 10^{-5} - 3.729 \times 10^{-4}$	0.002

*Predictor variables adjusted according to family history of obesity. CCI: conicity index; WHR: waist-hip ratio; FMI₁: fat mass distribution index 1; FMI₂: fat mass distribution index 2; IL: interleukin; TNF- α : tumor necrosis factor alpha.

of pro-inflammatory cytokines such as TNF- α , IL-6 and IL-12 in monocytes and macrophages. Therefore, there is a relationship between weight accumulation and these inflammatory indexes. Furthermore, a correlation was identified between adiposity indexes and cytokines: increased IL-12, IL-6 and TNF- α levels are associated with increased CCI, WHR, FMI₁ and FMI₂ indexes (Table V).

CCI assesses the tendency towards developing cardiovascular and metabolic diseases, allowing direct comparisons of abdominal adiposity between individuals (38).

FMI₁ and FMI₂ play a role in defining metabolic syndrome and abnormalities in the distribution of body fat (lipodystrophy) (39). Changes in the distribution of body fat can be accompanied by changes in blood glucose, enhancing any predisposition to develop cardiovascular diseases and diabetes. Although these changes are prevalent in HIV-positive patients, they are unknown in healthy individuals. However, it is inferred that they arise from a sedentary lifestyle and consumption of hypercaloric foods (40). From this perspective, an assessment of FMI₁ and FMI₂ is useful for diagnosing and treating clinical obesity, as well as for identifying individuals at potential risk for developing risk factors related to obesity (39).

In the light of the foregoing, the models for multiple linear regression analysis between body adiposity indexes and cytokines in the current study (Table V) show that, as cytokines increase, so do these indexes. The calculation of these indexes is based on anthropometric measures, which are easier to obtain and have a lower cost as compared to cytokine assessment, since its application in clinical practice will lead to lower healthcare expenditures.

It is noteworthy that, when interpreting the data from this study, it is appropriate to consider certain research limitations. The key one is its cross-sectional design. Thus, the associations presented

between independent factors and outcome variables do not necessarily represent causal relations, for this is the baseline of a longitudinal study. Therefore, the data shown can support further studies.

CONCLUSION

Pro-inflammatory cytokines in the current study were associated with an increase in adipose indexes. Obesity, especially morbid and visceral obesity, represents an inflammatory state, but its negative consequences are vitally important when it comes to cardiovascular disease. Therefore, these indexes may become a feasible strategy for clinical practice in order to identify propensity to inflammatory disorders.

REFERENCES

- Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* 2014;384(9945):766–81. DOI: 10.1016/S0140-6736(14)60460-8
- Casanello P, Krause BJ, Castro-Rodríguez JA, Uauy R. Epigenética y obesidad. *Rev Chil Pediatría* 2016;87(5):335–42. DOI: 10.1016/j.rchipe.2016.08.009
- Neeland IJ, Turer AT, Ayers CR, Powell-Wiley TM, Vega GL, Farzaneh-Far R, et al. Dysfunctional Adiposity and the Risk of Prediabetes and Type 2 Diabetes in Obese Adults. *JAMA* 2012;308(11):1150. DOI: 10.1001/2012.jama.11132
- Miazgowski T, Krzyżanowska-Świniarska B, Dziwura-Ogonowska J, Widecka K. The associations between cardiometabolic risk factors and visceral fat measured by a new dual-energy X-ray absorptiometry-derived method in lean healthy Caucasian women. *Endocrine* 2014;47(2):500–5. DOI: 10.1007/s12020-014-0180-7
- Hermesdorff HHM, Monteiro JBR. Gordura visceral, subcutânea ou intramuscular: onde está o problema? *Arq Bras Endocrinol Metabol* 2004;48(6):803–11. DOI: 10.1590/S0004-27302004000600005
- Bertin E, Marcus C, Ruiz J-C, Eschard J-P, Leutenegger M. Measurement of visceral adipose tissue by DXA combined with anthropometry in obese humans. *Int J Obes. Nature Publishing Group* 2000;24(3):263–70. DOI: 10.1038/sj.ijo.0801121

7. Direk K, Cecelja M, Astle W, Chowienczyk P, Spector TD, Falchi M, et al. The relationship between DXA-based and anthropometric measures of visceral fat and morbidity in women. *BMC Cardiovasc Disord* 2013;13(1):25. DOI: 10.1186/1471-2261-13-25
8. Amato MC, Giordano C, Galia M, Criscimanna A, Vitabile S, Midiri M, et al. Visceral Adiposity Index: A reliable indicator of visceral fat function associated with cardiometabolic risk. *Diabetes Care* 2010;33(4):920-2. DOI: 10.2337/dc09-1825
9. Grim M, Hertz B, Petosa R. Impact Evaluation of a Pilot Web-Based Intervention to Increase Physical Activity. *Am J Heal Promot* 2011;25(4):227-31. DOI: 10.4278/ajhp.081216-ARB-307
10. Haase A, Steptoe A, Sallis JF, Wardle J. Leisure-time physical activity in university students from 23 countries: Associations with health beliefs, risk awareness, and national economic development. *Prev Med (Baltim)* 2004;39(1):182-90. DOI: 10.1016/j.ypmed.2004.01.028
11. Irwin JD. Prevalence of university students' sufficient physical activity: a systematic review. *Percept Mot Skills* 2004;98(3 Pt 1):927-43. DOI: 10.2466/pms.98.3.927-943
12. Plotnikoff RC, Costigan S a, Williams RL, Hutchesson MJ, Kennedy SG, Robards SL, et al. Effectiveness of interventions targeting physical activity, nutrition and healthy weight for university and college students: a systematic review and meta-analysis. *Int J Behav Nutr Phys Act* 2015;12(1):45. DOI: 10.1186/s12966-015-0203-7
13. Correa-Rodríguez M, Ramírez-Vélez R, Correa-Bautista J, Castellanos-Vega R, Arias-Coronel F, González-Ruiz K, et al. Association of Muscular Fitness and Body Fatness with Cardiometabolic Risk Factors: The FUPRECOL Study. *Nutrients* 2018;10(11):1742. DOI: 10.3390/nu10111742
14. Shoelson SE, Herrero L, Naaz A. Obesity, Inflammation, and Insulin Resistance. *Gastroenterology* 2007;132(6):2169-80. DOI: 10.1053/j.gastro.2007.03.059
15. Prestes J, Donatto FF, Dias R, Frollini AB, Cavaglieri CR. Papel Da Interleucina-6 Como Um Sinalizador Em Diferentes Tecidos Durante O Exercício Físico. *Fit Perform J* 2006;5(6):348-53. DOI: 10.3900/fpj.5.6.348.p
16. Calder PC, Ahluwalia N, Albers R, Bosco N, Bourdet-Sicard R, Haller D, et al. A Consideration of Biomarkers to be Used for Evaluation of Inflammation in Human Nutritional Studies. *Br J Nutr. Cambridge University Press* 2013;109(S1):S1-34. DOI: 10.1017/S0007114512005119
17. Matsudo S, Araújo T, Matsudo V, Andrade D, Andrade E, Oliveira LC, et al. International Physical Activity Questionnaire (Ipaq): Validity and Reproducibility Study in Brazil. *Rev Bras Atividade Física Saúde* 2012;6(2):5-18. DOI: 10.12820/rbafs.v.6n2p5-18
18. Bergman RN, Stefanovski D, Buchanan TA, Sumner AE, Reynolds JC, Sebring NG, et al. A Better Index of Body Adiposity. *Obesity* 2011;19(5):1083-9. DOI: 10.1038/oby.2011.38
19. Vasques ACJ, Priore SE, Rosado LEFF de L, Franceschini S do CC. The use of anthropometric measures to assess visceral fat accumulation. *Rev Nutr* 2010;23(1):107-18. DOI: 10.1590/S1415-52732010000100012
20. World Health Organization (WHO). Diet, nutrition and the prevention of chronic diseases. Geneva: 2003; 2003. DOI: ISBN 92 4 120916 X ISSN 0512-3054 (NLM classification: QU 145)
21. Ashwell M, Gibson S. A proposal for a primary screening tool: "Keep your waist circumference to less than half your height". *BMC Med* 2014;12(1):207. DOI: 10.1186/s12916-014-0207-1
22. Morgan E, Varro R, Sepulveda H, Ember JA, Apgar J, Wilson J, et al. Cytometric bead array: a multiplexed assay platform with applications in various areas of biology. *Clin Immunol* 2004;110(3):252-66. DOI: 10.1016/j.clim.2003.11.017
23. World Health Organization. Global status report on noncommunicable diseases. World Health Organization; 2014. DOI: ISBN 9789241564854
24. Bogl LH, Kaye SM, Rämö JT, Kangas AJ, Soininen P, Hakkarainen A, et al. Abdominal obesity and circulating metabolites: A twin study approach. *Metabolism. Elsevier Inc* 2016;65(3):111-21. DOI: 10.1016/j.metabol.2015.10.027
25. Seyed-Sadjadi N, Berg J, Bilgin AA, Grant R. Visceral fat mass: Is it the link between uric acid and diabetes risk? *Lipids Health Dis* 2017;16(1):1-9. DOI: 10.1186/s12944-017-0532-4
26. Guglielmi V, Sbraccia P. Obesity phenotypes: Depot-differences in adipose tissue and their clinical implications. *Eat Weight Disord. Springer International Publishing* 2018;23(1):3-14. DOI: 10.1007/s40519-017-0467-9
27. Okosun IS, Seale JP, Lyn R. Commingling effect of gynoid and android fat patterns on cardiometabolic dysregulation in normal weight American adults. *Nutr Diabetes* 2015;5(5):e155-e155. DOI: 10.1038/nutd.2015.5
28. Chang S-H, Beason TS, Hunleth JM, Colditz GA. A systematic review of body fat distribution and mortality in older people. *Maturitas* 2012;72(3):175-91. DOI: 10.1016/j.maturitas.2012.04.004
29. Vella CA, Allison MA, Cushman M, Jenny NS, Miles MP, Larsen B, et al. Physical Activity and Adiposity-related Inflammation: The MESA. *Med Sci Sports Exerc* 2017;49(5):915-21. DOI: 10.1249/MSS.0000000000001179
30. Burghardt RD, Kazim MA, Rütther W, Niemeier A, Strahl A. The impact of physical activity on serum levels of inflammatory markers in rheumatoid arthritis: a systematic literature review. *Rheumatol Int. Springer Berlin Heidelberg* 2019;39(5):793-804. DOI: 10.1007/s00296-019-04284-x
31. Jeong S-K, Nam H-S, Son M-H, Son E-J, Cho K-H. Interactive Effect of Obesity Indexes on Cognition. *Dement Geriatr Cogn Disord* 2005;19(2-3):91-6. DOI: 10.1159/000082659
32. Hulsmans M, Van Dooren E, Mathieu C, Holvoet P. Decrease of miR-146b-5p in Monocytes during Obesity Is Associated with Loss of the Anti-Inflammatory but Not Insulin Signaling Action of Adiponectin. *PLoS One* 2012;7(2):e32794. DOI: 10.1371/journal.pone.0032794
33. Da Silva NI, Sobrinho HM da R, Blanch GT, Cruvinel WM, Gomes CM. Adipocinas e sua relação com a obesidade. *Rev EVS - Rev Ciências Ambient e Saúde* 2019;46(1). DOI: 10.18224/evs.v46i1.7179
34. El-Wakkad A, Hassan NE-M, Sibaii H, El-Zayat SR. Proinflammatory, anti-inflammatory cytokines and adiponectin in students with central obesity. *Cytokine* 2013;61(2):682-7. DOI: 10.1016/j.cyto.2012.11.010
35. Hunter CA. New IL-12-family members: IL-23 and IL-27, cytokines with divergent functions. *Nat Rev Immunol* 2005;5(7):521-31. DOI: 10.1038/nri1648
36. Da Silveira MR, Frollini AB, Verlengia R, Cavaglieri CR. Correlação entre obesidade, adipocinas e sistema imunológico. *Rev Bras Cineantropometria e Desempenho Hum* 2009;11(4):466-72. DOI: 10.5007/1980-0037.2009v11n4p466
37. FANTUZZI G. Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol* 2005;115(5):911-9. DOI: 10.1016/j.jaci.2005.02.023
38. Valdez R, Seidell JC, Ahn Yi, Weiss KM. A new index of abdominal adiposity as an indicator of risk for cardiovascular disease. A cross-population study. Vol. 17, *International journal of obesity and related metabolic disorders: journal of the International Association for the Study of Obesity*; 1993. p. 77-82.
39. Kelly TL, Wilson KE, Heymsfield SB. Dual Energy X-Ray Absorptiometry Body Composition Reference Values from NHANES. Vella A, directeur. *PLoS One* 2009;4(9):e7038. DOI: 10.1371/journal.pone.0007038
40. Viraben R, Aquilina C. Indinavir-associated lipodystrophy. *AIDS* 1998;12(6):F37-9. DOI: 10.1097/00002030-199806000-00001