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Inflammatory cytokines and non-alcoholic fatty liver disease (NAFLD) in obese children and adolescents

Citocinas inflamatorias y enfermedades hepáticas grasas no alcohólicas (EHGNA) en niños y adolescentes obesos

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ABSTRACT

Introduction: Non-alcoholic fatty liver disease is characterized by the intrahepatic deposition of fat. It is the most prevalent liver disease in the world, affecting obese

children and adolescents. Its pathophysiology is not fully understood, although it is often related to insulin resistance. This in turn would be due to an inflammatory condition common to obesity. Thus, the objective of this study was to describe the behavior of proinflammatory cytokines in obese children and adolescents, with and without non-alcoholic fatty liver disease.

Method: A fasting venous blood sample was obtained of consecutive 90 obese individuals aged 8-18 years, of both sexes, for laboratory determinations of glycaemia, basal insulin and homeostasis model assessment insulin-resistance index, and the inflammatory markers tumor necrosis factor-alpha (TNF- α), interleukins 2 and 6 (IL-2 and IL-6), interferon-gamma and high sensitive C-reactive protein. The clinical evaluation included weight, height and waist circumference. We used the body mass index/age indicator for the severity of overweight assessment. The degrees of steatosis were determined by ultrasonography. Quantitative and qualitative variables were respectively expressed by measures of central tendency/dispersion and simple/relative frequency, using Statistical Program for Social Sciences, version 20.0. A p-value < 0.05 was considered as significant.

Results: A total of 90 individuals were studied, with a mean age of 11.98 (2.72) years, of which 48 (53%) were male. The body mass index (BMI) for age (BMI/i) and sex (z-score) classified 38 (42.2%) participants as obese and 52 (57.7%) as severe obese; Hepatic steatosis was identified in 56 (62.2%) participants and approximately 90% of them presented grade I steatosis. The inflammatory markers TNF- α , and C-reactive protein were increased in the studied sample and correlated in a positive and statistically significant way with the index of body mass/age and sex.

Conclusion: Hepatic steatosis was prevalent in the group of children and adolescents studied, but was not related to obesity degrees.

Key words: Non-alcoholic fatty liver disease. Obesity. Fatty liver. Children. Adolescents.

RESUMEN

Introducción: la enfermedad hepática grasa no alcohólica se caracteriza por la deposición intrahepática de grasa. Es la enfermedad hepática más prevalente en el mundo y afecta a niños y adolescentes obesos. Su fisiopatología no se entiende

completamente, aunque a menudo se relaciona con la resistencia a la insulina. Esto, a su vez, sería debido a una condición inflamatoria común a la obesidad. Así, el objetivo de este estudio fue describir el comportamiento de las citocinas proinflamatorias en niños y adolescentes obesos, con y sin enfermedad hepática grasa no alcohólica.

Método: se obtuvo una muestra de sangre venosa en ayuno de 90 individuos obesos consecutivos, de 8 a 18 años, de ambos sexos, para las determinaciones de laboratorio de glucosa, insulina basal, resistencia a la insulina (*homeostasis model assessment insulin-resistance index*) y marcadores inflamatorios como TNF-alfa, interleucinas 2 y 6, interferón-gamma y proteína C-reactiva ultrasensible. La evaluación clínica incluyó peso, altura y circunferencia de la cintura. El indicador del índice de masa corporal para la edad (IMC/e) y sexo (*z-score*) evaluó la gravedad del exceso de peso. Los grados de esteatosis se determinaron por ecografía. Las variables cuantitativas y cualitativas se expresaron respectivamente mediante tendencia/dispersión central y frecuencia simple/relativa, utilizando el Programa Estadístico para Ciencias Sociales, versión 20.0. Se consideró significativo un valor $p < 0,05$.

Resultados: fueron estudiados 90 pacientes con una media de edad de 11,98 (2,72) años, de los cuales 48 (53%) eran varones. Según IMC/e y sexo, 38 (42.2%) participantes fueron clasificados como obesos y 52 (57,7%), como obesos graves. La esteatosis hepática fue identificada en 56 (62,2%) participantes y aproximadamente el 90% de ellos presentaron esteatosis grado I. Los marcadores inflamatorios TNF-alfa y proteína C-reactiva estaban aumentados en la muestra estudiada y se correlacionaron de forma positiva y estadísticamente significativa con el IMC/e y sexo.

Conclusión: la esteatosis hepática fue predominante en el grupo de niños y adolescentes estudiados, pero no se relacionó con los grados de obesidad.

Palabras clave: Enfermedad del hígado graso no alcohólico. Obesidad. Hígado graso. Niño. Adolescente.

INTRODUCTION

Obesity is a chronic disease that affects the population throughout the world, being more prevalent than malnutrition and infectious diseases (1). Studies reveal that obesity affects not only adults but is also verified in children and adolescents (2,3),

when related metabolic alterations may have its onset (4). Adipose tissue is characterized, in particular, by its complexity. It would originally be related to energy source and thermogenesis, depending on its differentiation in white adipose tissue (WAT) or brown adipose tissue (BAT), respectively. However, other functions have been attributed to adipose tissue, resulting from its active involvement in immune, hormonal and metabolic processes through the synthesis and release of substances called adipokines (5). Adipose tissue is recognized as an endocrine organ for its secretory and synthesizing function of leptin, estrogens, angiotensinogen and adiponectin, by the ability to establish connections with the central nervous system, in addition to the release of acute-phase proteins and pro-inflammatory cytokines such as tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6) and insulin-like growth factor 1 (IGF-1) (6). TNF- α and IL-6 cytokines are related to obesity and reduced insulin sensitivity (7). Insulin resistance would lead to increased synthesis and storage of triglycerides in hepatocytes, and thus play an important role in the pathogenesis of non-alcoholic fatty liver disease (NAFLD), a disease with a high potential for cirrhosis and/or hepatocellular carcinoma (8). As a consequence, this population could develop other metabolic alterations such as dyslipidemias, hypertension and type 2 diabetes mellitus (T2DM), secondary to a supposed inflammatory state intrinsic to obesity that would respond by the expression of these comorbidities, according to studies described in adults (9). Considering these aspects, the purpose of this study was to describe the behavior of proinflammatory cytokines in obese children and adolescents with and without non-alcoholic fatty liver disease (NAFLD).

METHODS

A cross-sectional study involving 90 obese children and adolescents of both sexes, aged between eight and 18 years, consecutively evaluated at the Pediatric Nutrology Service of the Federal University of Bahia from January to December 2015, was carried out, featuring a convenience sample. Patients with conditions predisposing to overweight such as Cushing's syndrome, growth hormone deficiency, hypothyroidism, and syndromic obesity were excluded. There was no report of consumption of alcoholic beverages among participants. The body mass index (BMI) was calculated by dividing the weight (kg) by the square of the height (m) (kg/m^2). The z-score of the BMI

indicator/age and gender was used to classify the participants' anthropometric status as obese (BMI/i > +2) and severe obese (BMI/i > +3) (10). Waist circumference was obtained with a soft and inelastic tape, at the midpoint between the last rib and the anterior iliac crest (11). GE LOGIC P6 ultrasound with a convex transducer of 2 to 5 MHz was used for the diagnosis and classification of steatosis in grades I, II and III, according to the alteration of echogenicity, and the identification of intrahepatic vessels and diaphragm, according to Hamaguchi et al. (12). The imaging examinations were performed at the research institution and by the same professional. This study was approved by the Ethics Committee from the Federal University of Bahia, Brazil. A children verbal assent and a written consent for participation were obtained through their legal representatives.

Laboratory analyses

A fasting venous blood sample was taken of all participants for laboratory determinations. The enzymes alanine transaminase (ALT), aspartate transaminase (AST) and gamma-glutamyl transpeptidase (gamma-GT) were measured by the calorimetric kinetic method and expressed in U/l. Altered fasting plasma glucose ≥ 100 mg/dl was assessed by the hexokinase method (Wiener lab.) (13). Insulin levels were evaluated by enzyme linked immunoenzymatic assay (ELISA). Any values greater than 15.0 μ U/ml were considered as abnormal (14). The homeostasis model assessment-insulin resistance (HOMA-IR) index = fasting glucose (mg/dl) x fasting insulin (μ IU ml⁻¹)/405 was used to identify insulin resistance (15). We used the cut-offs adjusted by gender and age according to Almeida et al., summing the mean value to two standard deviations (16). The high sensitivity C-reactive protein (hsCRP) was determined by immunoturbidimetric method. The interleukin-2 (IL-2), interleukin-6 (IL-6), interferon-gamma (IFN- γ) and tumor necrosis factor- α (TNF- α) were determined by ELISA DuoSet R&D Systems.

Statistical analyses

The Statistical Program for Social Sciences (SPSS), version 20.0, was used for statistical analyses. Quantitative and qualitative data were respectively expressed by measures of central tendency/dispersion and simple/relative frequency. Continuous variables

were tested for normality of distribution by Kolmogorov-Smirnov test. The differences for these variables were analyzed using the Mann-Whitney U-test or Student's t-test, according to the distribution. The Chi-squared test compared the frequency of individuals in each category. A p-value < 0.05 was considered to be significant.

RESULTS

Ninety individuals were studied, with a mean age of 11.98 (2.72) years, of which 48 (53%) were male. The BMI for age (BMI/i) and sex (z-score) classified 38 (42.2%) participants as obese and 52 (57.7%) as severe obese, being 3.14 (0.86) years the mean (SD) and 2.02 and 8.52 the minimum and maximum values of BMI/i, respectively. Hepatic steatosis was identified in 56 (62.2%) participants, being more frequent in boys than in girls (58.9 *versus* 41.1%, $p > 0.05$). Among these, 50 (89.2%) presented grade I steatosis, five (8.92%) presented grade II and only one (1.78%), grade III steatosis. No case of steatohepatitis (NASH) was identified, considering that the levels of transaminases were within the reference values. Blood pressure levels were measured and found within normal limits. Table 1 shows the biochemical parameters and the inflammatory markers analyzed considering the presence of NAFLD.

The inflammatory profile of all study participants considering obesity degrees is shown in table 2. Table 3 shows the clinical and laboratory characteristics of obese and severe obese patients, considering the presence of NAFLD. Table 4 shows that there was a positive and statistically significant correlation between HOMA-IR, insulin and CRP, different from that of IFN- γ , IL-6, and IL-2. BMI/age also influenced the behavior of TNF- α and CRP.

DISCUSSION

The relevance of this study is due to the fact that the altered findings were from a sample of 90 clinically asymptomatic obese individuals. Hepatic steatosis was prevalent, confirming literature data indicating NAFLD as one of the adverse consequences of obesity, in addition to hyperlipidemia, hypertension and type 2 diabetes mellitus (T2DM) (17). Grade I steatosis was identified in approximately 90% of the 56 children and adolescents diagnosed with NAFLD, reflecting a process of early and incipient installation of the spectrum involving the disease.

In table 1, groups with and without liver disease are compared. We observed that individuals with NAFLD did not present significant differences in the analyzed parameters. However, in table 2, inflammatory markers such as adipokine TNF- α were elevated in severe obese individuals, whereas CRP, an acute phase protein of inflammation secreted by the liver when stimulated by IL-6, was increased in obese individuals, suggesting the presence of an inflammatory state underlying obesity.

In addition, in table 3, the 56 individuals diagnosed with hepatic steatosis were analyzed considering the obesity degree, and statistically significant changes were found. In severe obese patients, waist circumference was higher, probably due to the central distribution pattern of adiposity secondary to insulin resistance. Likewise, severe obese individuals presented the highest rates of GGT. In their study, Fishbein et al. (18) examined the relationship of transaminases with the severity of steatosis and concluded that major changes are expected in severe cases of fatty liver, as opposed to our findings, given the prevalence of steatosis in the early stage has been similar between obese and severe obese. No association between the inflammatory profile and serum GGT values was observed.

As previously mentioned, insulin resistance (IR) is the main determinant of the pathophysiology of NAFLD (8). We observed that the IR indicators (HOMA-IR and serum insulin) were higher in obese patients with NAFLD, but did not differ between patients with and without liver disease. In parallel, TNF- α , an inflammatory cytokine associated with obesity and insulin resistance, showed similar behavior. TNF- α plays a crucial role in the determination of insulin resistance (19), stimulates the synthesis of other cytokines and has direct action in the liver by the increase of lipogenesis by upregulating catalyzing enzymes (20). In addition, it suppresses the expression of genes involved in the uptake of glucose and fatty acid metabolism favoring its accumulation (21). Finally, the inflammatory cascade triggered by TNF- α would be related to the progression of NAFL to NASH (22). Thus, it can be pointed out that these children and adolescents with NAFLD and severe obesity should be monitored more frequently in the future, since at the time of this study, from the laboratory point of view, there were no indications of hepatitis.

Jovinge et al. (23) report an increase in TNF- α in male subjects related to atherosclerotic disease, although this has not been verified in relation to insulin

resistance. However, other studies (24,25) performed in adults describe a significant correlation between TNF- α , serum insulin and body mass index. In this regard, we evaluated the behavior of TNF- α in cases where HOMA-IR and serum insulin were normal or altered and no statistically significant differences were detected, although this behavior was confirmed when the BMI/age was considered. In addition, a correlation analysis involving the 56 children and adolescents diagnosed with NAFLD was performed. A positive and statistically significant correlation was found between HOMA-IR and serum insulin and CRP, an acute phase protein of inflammation, which has serum levels directly proportional to body mass index and therefore associated with obesity (26). Inversely, the IL-6, an inflammatory cytokine associated with obesity and insulin resistance, secreted by T and B lymphocytes, endothelium, fibroblasts and macrophages, showed a negative and significant correlation with IL2, a cytokine that participates in the immune response, secreted by T lymphocytes, suggesting a modulatory effect between cytokines (27). Thus, in our study we may suggest that changes in inflammatory markers may have influenced glycemic metabolic indicators with respect to insulin resistance and NAFLD in subjects with severe obesity.

Most likely, limitations of the study refer to its cross-sectional design, which because of its characteristics does not make it possible to establish a cause and effect relationship. Likewise, individual immune variations, the complexity involving the characterization of the inflammatory status and the verification of the cytokine dosages in a single moment probably may have compromised the verification of the inflammatory framework intrinsic to obesity. We also emphasize limitations involving the imaging method (ultrasound) used for the diagnosis of NAFLD, which does not allow determining the severity of the inflammation with respect to the presence of steatohepatitis or fibrosis, which would only be allowed through liver biopsy, considered as gold standard.

Studies (28,29) are carried out in an attempt to elucidate an alleged low-grade inflammatory process underlying obesity, which would account for the expression of associated comorbidities, such as type 2 diabetes mellitus, systemic arterial hypertension, dyslipidemias and fatty liver. However, understanding is neither complete nor sufficient to define whether obesity will lead to an inflammatory picture or whether inflammation will lead to obesity. In our study, no differences in the profile

of inflammatory cytokines between patients with and without NALFD were observed. However, the severely obese had higher median levels of TNF- α and IL-6, although the latter was not significant ($p = 0.059$), showing, however, a tendency for association. Although not statistically significant, the proportion of serious obese children and adolescents without NALFD was significantly higher than those with this change (52.9 vs 35.7%), suggesting that obesity severity is a confounder of this relationship. In fact, we found that the TNF- α profile was only higher among the patients, and it was significant among the obese patients when compared to obese patients. A possible explanation for this finding corresponds to the greater mass of adipocytes among the severe obese, confirmed by the larger waist circumferences presented by this group. In fact, abdominal adiposity is associated with increased insulin resistance and hyperinsulinism. Thus, these patients would carry a greater amount of pro-inflammatory cytokines. In addition, considering only the severely obese, we observed that this condition was associated with the most suggestive parameter of NAFLD, which is insulin resistance, variable related to the entire spectrum of the disease, from the emerging steatosis to the progression of the disease to cirrhosis and hepatocarcinoma (30).

Thus, our study suggests that obesity is an inflammatory condition in isolation. In some cases, comorbidities such as NAFLD can be expressed. Thus, we suggest the investigation of this population in view of the presumption of an incipient inflammatory condition since the onset of obesity.

Finally, this study intends to establish a theoretical reference, considering that the search in the literature resulted in a greater number of articles of revision when compared to the original articles.

REFERENCES

1. WHO. Obesity preventing and managing the global epidemic. Report of WHO Consultation on Obesity. Geneva: World Health Organization; 1998.
2. Ogden CL, Flegal KM, Carroll MID, Johnson CL. Prevalence and trends in obesity among US children and adolescents 1999-2000. JAMA 2002;288:1728-32.3.
3. Costa RF, Cintra IP, Fisberg M. Prevalence of overweight and obesity in school children of Santos city, Brazil. Arq Bras Endocrinol Metab 2006;50(1):60-7.

4. Harrell JS, Jessup A, Greene N. Changing our future: Obesity and the metabolic syndrome in children and adolescents. *J Cardiovasc Nurs* 2006;21:322-30.
5. Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab* 2004;89(6):2548-56.
6. Wajchenberg BL. Tecido adiposo como glândula endócrina. *Arq Bras Endocrinol Metab* 2000;44/1:1320.
7. Fonseca-Alaniz MH, Takada J, Alonso-Vale MI, Lima FB. The adipose tissue as a regulatory center of the metabolism. *Arq Bras Endocrinol Metab* 2006;50:216-29.
8. Carvalheira JBC, Saad MJA. Insulin resistance/hyperinsulinemia associated diseases not included in the metabolic syndrome. *Arq Bras Endocrinol Metab* 2006;50(2):360-7.
9. Codoner-Franch P, Valls-Belles V, Arilla-Codoner A, Alonso-Iglesias E. Oxidant mechanisms in childhood obesity: The link between inflammation and oxidative stress. *Transl Res* 2011;158(6):369-84.
10. Onis M, Onyango AW, Borghi E, Siyam A, Nishida C, Siekmann J. Development of a WHO growth reference of school aged children and adolescents. *Bulletin of the WHO* 2007;660-7.
11. WHO. Obesity: Preventing and managing the global epidemic. Geneva: World Health Organization; 2000.
12. Hamaguchi M, Kojima T, Itoh Y, Harano Y, Fujii K, Nakajima T, et al. The severity of ultrasonographic findings in nonalcoholic fatty liver disease reflects the metabolic syndrome and visceral fat accumulation. *Am J Gastroenterol* 2007;102(2):2708-15.
13. American Diabetes Association. Diagnosis and classification of diabetes mellitus. *Diabetes Care* 2014;37(Suppl 1)36:S81-90.
14. Obesidade na infância e na adolescência - Manual de Orientação/Sociedade Brasileira de Pediatria. Departamento Científico de Nutrologia. 2nd ed. São Paulo: SBP; 2012. p. 142.
15. Geloneze B, Tambascia MA. Laboratorial evaluation and diagnosis of insulin resistance. *Arq Bras Endocrinol Metab* 2006;50(2):208-15.
16. Almeida CAN, Pinho AP, Ricco RG, Pepato MT, Brunetti IL. Determination of glycemia and insulinemia and the homeostasis model assessment (HOMA) in

schoolchildren and adolescents with normal body index. *J Pediatr (Rio J)* 2008;84:136-40.

17. Roberts EA. Pediatric nonalcoholic fatty liver disease (NAFLD): A “growing” problem? *J Hepatol* 2007;46:1133-42.

18. Fishbein MH, Miner M, Mogren C, Chalekson J. The spectrum of fatty liver in obese children and the relationship of serum aminotransferases to severity of steatosis. *J Pediatr Gastroenterol Nutr* 2003;36(1):54-61.

19. Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004;145:2273-82.

20. Chitturi S, Abeygunasekera S, Farrell GC, Holmes-Walker J, Hui JM, Fung C, et al. NASH and insulin resistance: Insulin hypersecretion and specific association with the insulin resistance syndrome. *Hepatology* 2002;35:373-9.

21. Ruan H, Miles PDG, Ladd CM, Ross K, Golub TR, Olefsky JM, et al. Profiling gene transcription in vivo reveals adipose tissue as an immediate target tumor necrosis factor- α : Implications for insulin resistance. *Diabetes* 2002;51:3176-88.

22. Peverill W, Powell LW, Skoien R. Evolving concepts in the pathogenesis of NASH: Beyond steatosis and inflammation. *Int J Mol Sci* 2014;15:8591-638.

23. Jovinge S, Hamsten A, Tornvall P, Proudler A, Bävrenhoim P, Ericsson C-G, et al. Evidence for a role of tumor necrosis factor-alpha in disturbance of triglyceride and glucose metabolism predisposing to coronary heart disease. *Metabolism* 1998;47:113-8.

24. Hotamisligil GS, Arner P, Caro JF, Atkinson RL, Spiegelman BM. Increased adipose tissue expression of tumor necrosis factor-alpha in human obesity and insulin resistance. *J Clin Invest* 1995;95:2409-15.

25. Kern PA, Sagnizadeh M, Ong JM, Bosch RJ, Deem R, Simsolo RB. The expression of tumor necrosis factor in human adipose tissue. Regulation by obesity, weight loss, and relationship to lipoprotein lipase. *Clin Invest* 1995;95:2111-29.

26. Trayhurn P, Wood IS. Adipokines: Inflammation and the pleiotropic role of white adipose tissue. *Br J Nutr* 2004;92(3):347-55.

27. Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 pathogenic T-cell population induces autoimmune inflammation. *J Exp Med* 200;201(2):233-40.
28. Caballero B. The global epidemic of obesity: An overview. *Epidemiol Rev* 2007;29:1-5.
29. Spielgeman BM, Flier JS. Obesity and the regulation of energy balance. *Cell* 2001;104:531-43.
30. Duvnjak M, Lerotic I, Barsic N, Tomasic V, Jukic LV, Velagie V. Pathogenesis and management issues for non-alcoholic fatty liver disease. *World J Gastroenterol* 2007;13:4539-50.

Table 1. Clinical and laboratory characteristics of 90 children and adolescents studied according to the presence of non-alcoholic fatty liver disease

| | NAFLD Yes (n = 56) | NAFLD No (n = 34) | p value |
|---------------------------|-----------------------|----------------------|---------|
| Age (years)** | 12.00 (10-14) | 11.00 (10-14) | 0.687 |
| Male gender (%) | 58.9 | 44.1 | 0.172 |
| BMI/age (z-score)* | 3.15 (0.61) | 3.13 (1.18) | 0.884 |
| Severe obese | 20 (35.7%) | 18 (52.9%) | 0.109 |
| Waist circumference (cm)* | 96.56 (12.67) | 92.82 (14.54) | 0.203 |
| ALT (UI)* | 26.09 (10.97) | 22.50 (6.17) | 0.084 |
| AST (UI)** | 23.00 (15.00-28.00) | 20.00 (12.00-28.00) | 0.237 |
| GGT (UI)** | 25.00 (20.00-35.00) | 22.00 (17.00-28.00) | 0.106 |
| Cholesterol (mg/dl)* | 166.82 (34.79) | 163.01 (34.58) | 0.617 |
| LDL (mg/dl)* | 97.50 (31.61) | 93.30 (33.80) | 0.563 |
| HDL (mg/dl)* | 42.32 (7.17) | 43.63 (9.69) | 0.468 |
| Triglycerides (mg/dl)** | 117 (82.23-144.50) | 95.35 (72.50-172.25) | 0.659 |
| Glycemia (mg/dl)* | 91.39 (9.96) | 87.64 (9.06) | 0.077 |
| HOMA-IR** | 2.54 (1.55-3.26) | 2.18 (1.36-3.89) | 0.600 |
| Insulin (mUI)** | 11.63 (7.37-15.22) | 10.12 (6.94-17.84) | 0.623 |
| hs-CRP (mg/l)** | 4.05 (1.97-7.28) | 3.88 (1.88-13.67) | 0.519 |

| | | | |
|-------------------------|---------------------|---------------------|-------|
| Interleukin-2 (pg/ml)** | 0.240 (0.097-0.425) | 0.192 (0.069-0.351) | 0.265 |
| Interleukin-6 (pg/ml)** | 0.287 (0.121-0.770) | 0.206 (0.058-0.658) | 0.222 |
| TNF- α (pg/ml)** | 0.269 (0.119-0.642) | 0.235 (0.101-0.453) | 0.268 |
| IFN γ (pg/ml)** | 0.352 (0.103-0.721) | 0.213 (0.075-0.535) | 0.144 |

BMI: Body mass index; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; GGT: Gamma glutamyl transpeptidase; HDL: High-density lipoprotein; LDL: Low-density lipoprotein; HOMA-IR: Homeostasis model assessment-insulin resistance; hs-CRP: High sensitivity C reactive protein; TNF- α : Tumor necrosis factor- α ; IFN- γ : Interferon gamma. *Values express as mean (SD). ** Values express as median and interquartile.

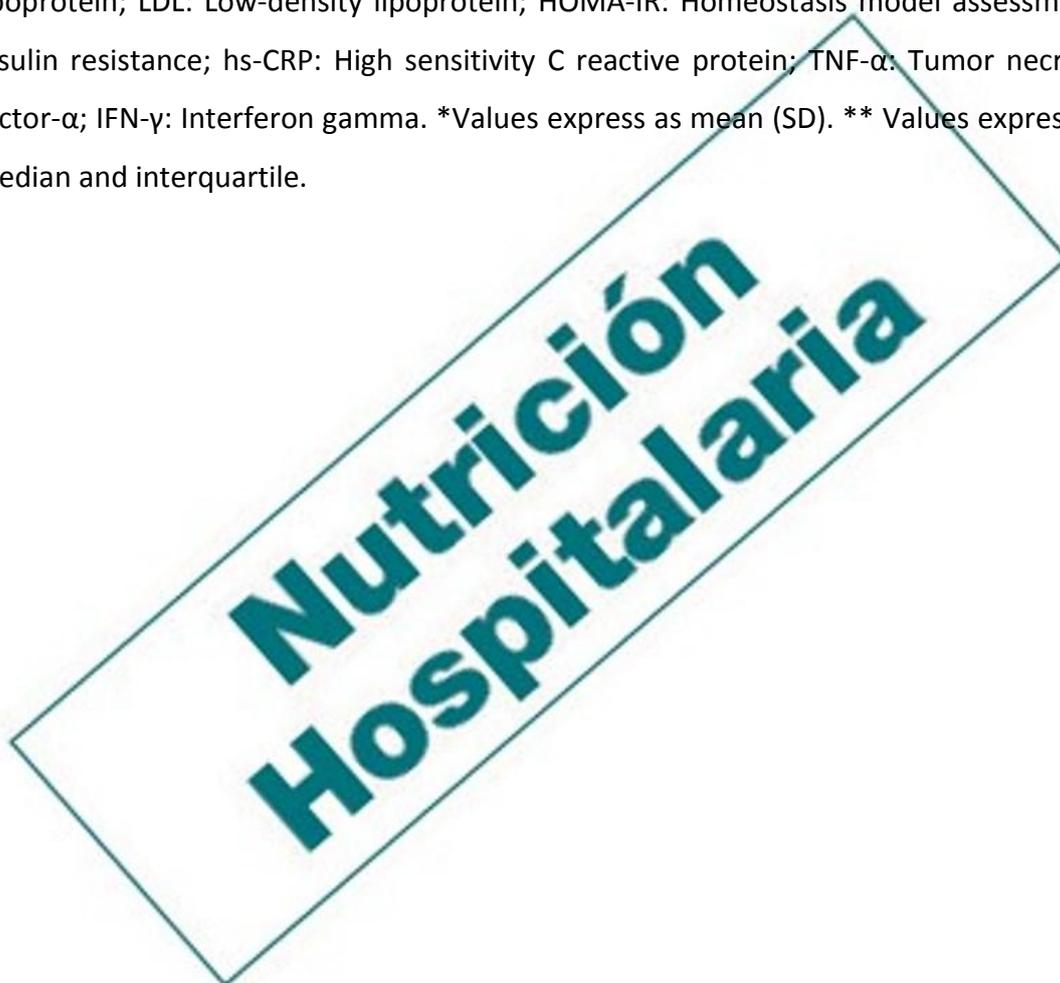


Table 2. Behavior of inflammatory markers of 90 children and adolescents studied, according to obesity degrees

| | Severe obese | Obese | p value |
|------------------------|---------------------|---------------------|---------|
| hs-CRP (mg/l)* | 2.52 (1.16-5.37) | 5.16 (2.94-9.07) | 0.009 |
| Interleukin-2 (pg/ml)* | 0.222 (0.104-0.456) | 0.208 (0.085-0.400) | 0.520 |
| Interleukin-6 (pg/ml)* | 0.515 (0.116-0.865) | 0.212 (0.091-0.559) | 0.088 |
| TNF- α (pg/ml)* | 0.315 (0.190-0.667) | 0.204 (0.084-0.428) | 0.024 |
| IFN- γ (pg/ml)* | 0.230 (0.072-0.693) | 0.281 (0.084-0.609) | 0.750 |

hs-CRP: High sensitivity C reactive protein; TNF- α : Tumor necrosis factor- α ; IFN- γ : Interferon gamma. *Values express as median and interquartiles.

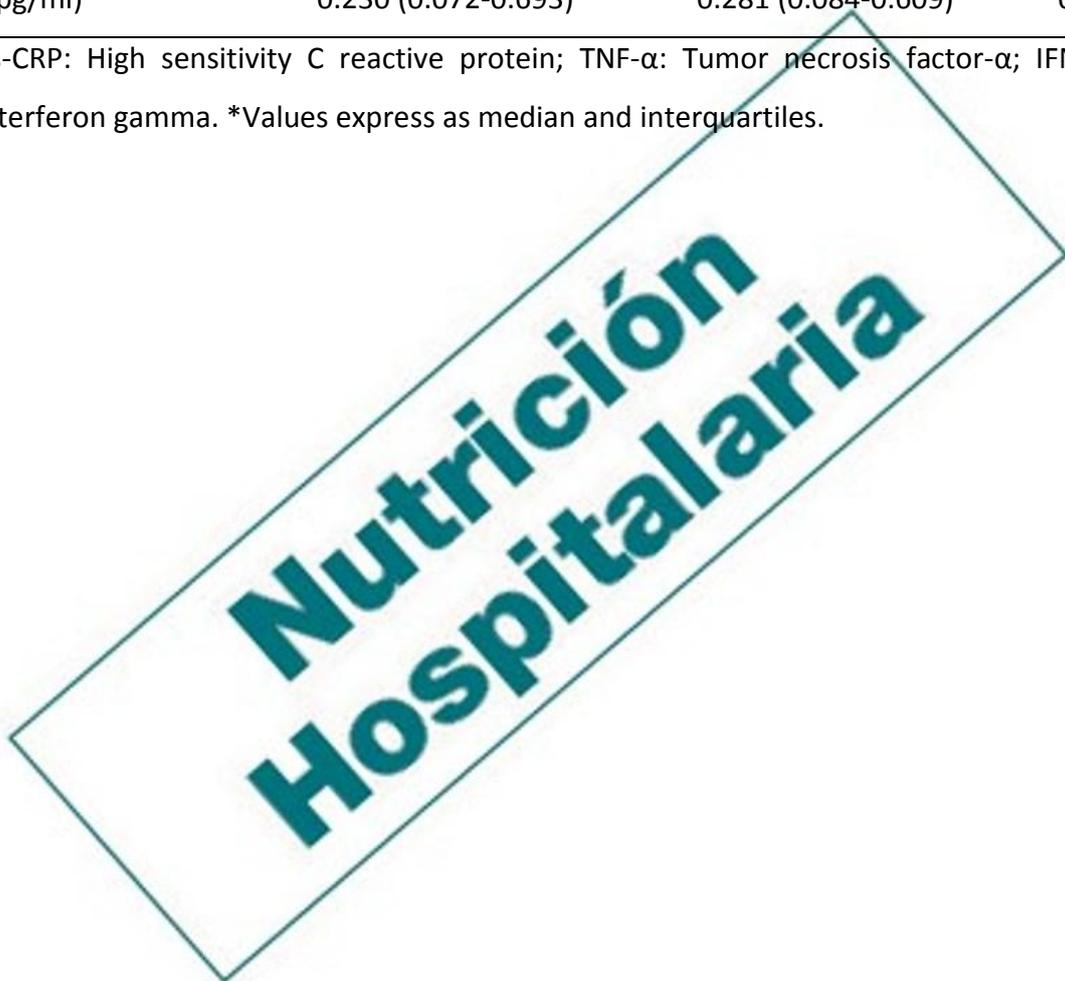


Table 3. Clinical and laboratory characteristics of 56 children and adolescents with non-alcoholic fatty liver disease according to the obesity degree

| | Severe obese (n = 20) | Obese (n = 36) | p value |
|---------------------------|--------------------------|---------------------|---------|
| Age (years)* | 13.30 (2.55) | 11.36 (2.58) | 0.009 |
| Gender (%) | 58.9% | 41.1% | 0.909 |
| BMI/age (z-score)* | 3.60 (0.87) | 2.52 (0.25) | 0.007 |
| Degrees of steatosis*: | | | 0.597 |
| - grade I | 18 (90.0%) | 32 (88.9%) | |
| - grade II | 2 (10.0%) | 3 (8.3%) | |
| - grade III | 0 | 1 (2.8%) | |
| Waist circumference (cm)* | 98.01 (13.63) | 93.67 (10.04) | 0.049 |
| ALT (UI)* | 26.60 (12.17) | 25.80 (10.40) | 0.800 |
| AST (UI)** | 21.00 (16.00-33.00) | 18.50 (12.00-28.00) | 0.782 |
| GGT (UI)** | 26.00 (22.00-38.00) | 20.00 (17.00-28.00) | 0.002 |
| Cholesterol (mg/dl)* | 173.42 (32.50) | 155.80 (36.47) | 0.069 |
| LDL (mg/dl)* | 103.55 (31.96) | 87.20 (33.96) | 0.066 |
| HDL (mg/dl)* | 47.97 (10.48) | 40.84 (9.67) | 0.220 |
| Triglycerides (mg/dl)* | 147.56 (86.75) | 129.99 (78.18) | 0.464 |
| Glycemia (mg/dl)* | 97.90 (9.45) | 88.09 (10.51) | 0.450 |
| HOMA-IR** | 2.56 (1.69-5.11) | 2.03 (1.20-2.87) | 0.009 |
| Insulin (mUI)** | 14.01 (8.89-21.90) | 8.66 (6.22-11.72) | 0.001 |
| hs-CRP (mg/l)** | 1.94 (0.80-2.28) | 1.30 (0.71-2.00) | 0.484 |
| Interleukin-2 (pg/ml)** | 0.279 (0.101-0.345) | 0.231 (0.094-0.365) | 0.430 |
| Interleukin-6 (pg/ml)** | 0.490 (0.109-0.754) | 0.320 (0.085-0.487) | 0.059 |
| TNF- α (pg/ml)** | 0.390 (0.187-0.695) | 0.301 (0.074-0.328) | 0.032 |
| IFN- γ (pg/ml)** | 0.340 (0.091-0.609) | 0.233 (0.098-0.687) | 0.670 |

BMI: Body mass index; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; GGT: Gamma glutamyl transpeptidase; LDL: Low-density lipoprotein; HDL: High-density lipoprotein; HOMA-IR: Homeostasis model assessment-insulin resistance; hs-CRP: High sensitivity C reactive protein; TNF- α : Tumor necrosis

factor- α ; IFN- γ : Interferon gamma. * Values express as mean (SD). ** Values express as median and interquartiles.



Table 4. Correlation analysis of inflammatory markers and clinical and laboratory parameters of 56 children and adolescents with non-alcoholic fatty liver disease

| | IL2 | IL6 | TNF- α | IFN- γ | High sensitivity PCR |
|--------------------------|------------------|------------------|---------------|---------------|----------------------|
| Age (years) | 0.066; 0.631 | -0.127; 0.072 | -0.095; 0.485 | -0.112; 0.410 | -0.182; 0.457 |
| Waist circumference (cm) | 0.061; 0.656 | -0.124; 0.362 | 0.073; 0.595 | -0.072; 0.600 | -0.134; 0.584 |
| BMI | -0.119; 0.264 | -0.078; 0.465 | 0.122; 0.251 | 0.011; 0.921 | 0.183; 0.084 |
| BMI/age | -0.131; 0.219 | -0.111; 0.297 | 0.272; 0.009 | 0.062; 0.562 | 0.285; 0.006 |
| ALT (UI) | -0.029; 0.834 | -0.058; 0.674 | 0.126; 0.359 | 0.012; 0.929 | 0.076; 0.756 |
| AST (UI) | -0.029; 0.832 | 0.016; 0.905 | 0.016; 0.905 | 0.235; 0.084 | -0.183; 0.454 |
| GGT (UI) | -0.029; 0.832 | -0.181; 0.186 | -0.094; 0.510 | 0.156; 0.273 | 0.090; 0.613 |
| Glycemia (mg/dl) | -0.237; 0.079 | -0.090; 0.512 | -0.134; 0.325 | 0.251; 0.062 | -0.275; 0.254 |
| HOMA-IR | 0.005; 0.969 | 0.045; 0.741 | -0.055; 0.685 | 0.109; 0.424 | 0.407; 0.034 |
| Insulin (mUI) | -0.083; 0.542 | 0.020; 0.886 | -0.036; 0.795 | 0.086; 0.527 | 0.484; 0.036 |
| hs-CRP (mg/l) | 0.078; 0.750 | -0.452; 0.050 | 0.356; 0.135 | -0.257; 0.295 | 1 |
| Interleukin-2 (pg/ml) | 1 | -0.305; 0.022 | -0.163; 0.230 | -0.377; 0.004 | 0.078; 0.750 |
| Interleukin-6 (pg/ml) | -0.305; 0.022 | 1 | -0.113; 0.407 | 0.128; 0.349 | -0.452; 0.052 |
| TNF- α (pg/ml) | -0.163; | -0.113; | 1 | -0.260; 0.053 | 0.356; 0.135 |

| | | | | | |
|-----------------------|-------------------------|-----------------|---------------|---|---------------|
| | 0.230 | 0.407 | | | |
| IFN- γ (pg/ml) | -0.377; <i>0.004</i> | 0.128; 0.349 | -0.260; 0.053 | 1 | -0.254; 0.295 |

BMI: Body mass index; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; GGT: Gamma glutamyl transpeptidase; HOMA-IR: Homeostasis model assessment-insulin resistance; hs-CRP: C reactive protein-high sensitivity; TNF- α : Tumor necrosis factor- α ; IFN- γ : Interferon gamma.

